

Metagenomics 101

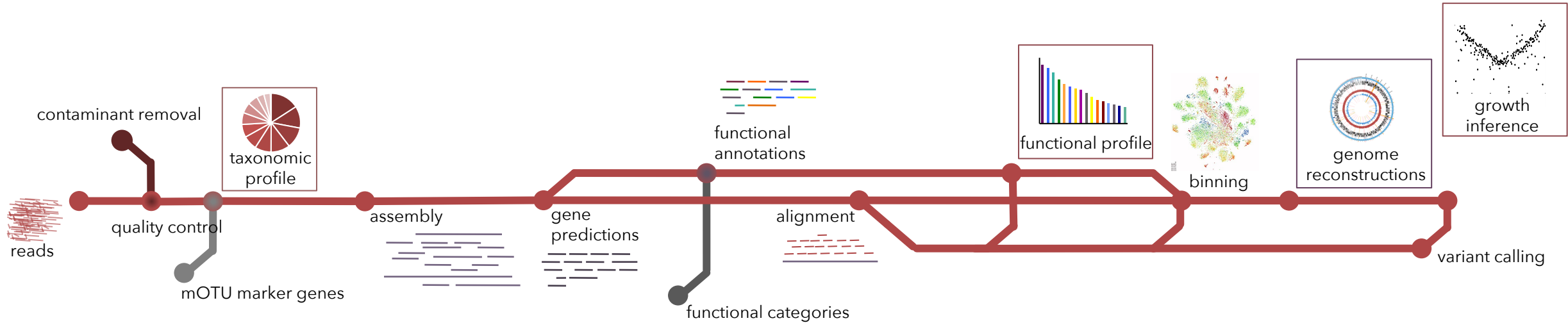
Session X: Metatranscriptomics

Anna Heintz-Buschart

April 2022



Metagenomics (+ other omics) pipeline

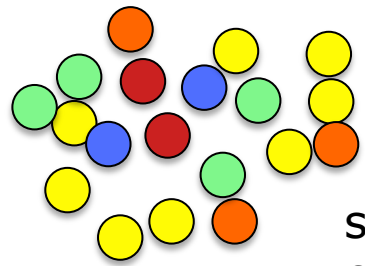


Today

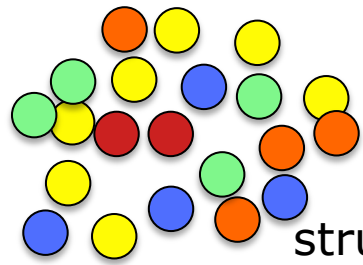
- Why metatranscriptomics?
- Challenges in metatranscriptomics
- Workflows for metatranscriptomics

Metaomics

rRNA amplicon studies:



structure of community A



structure of community B

metagenomics:



potential of community A



potential of community B

functional omics:

activity of community A

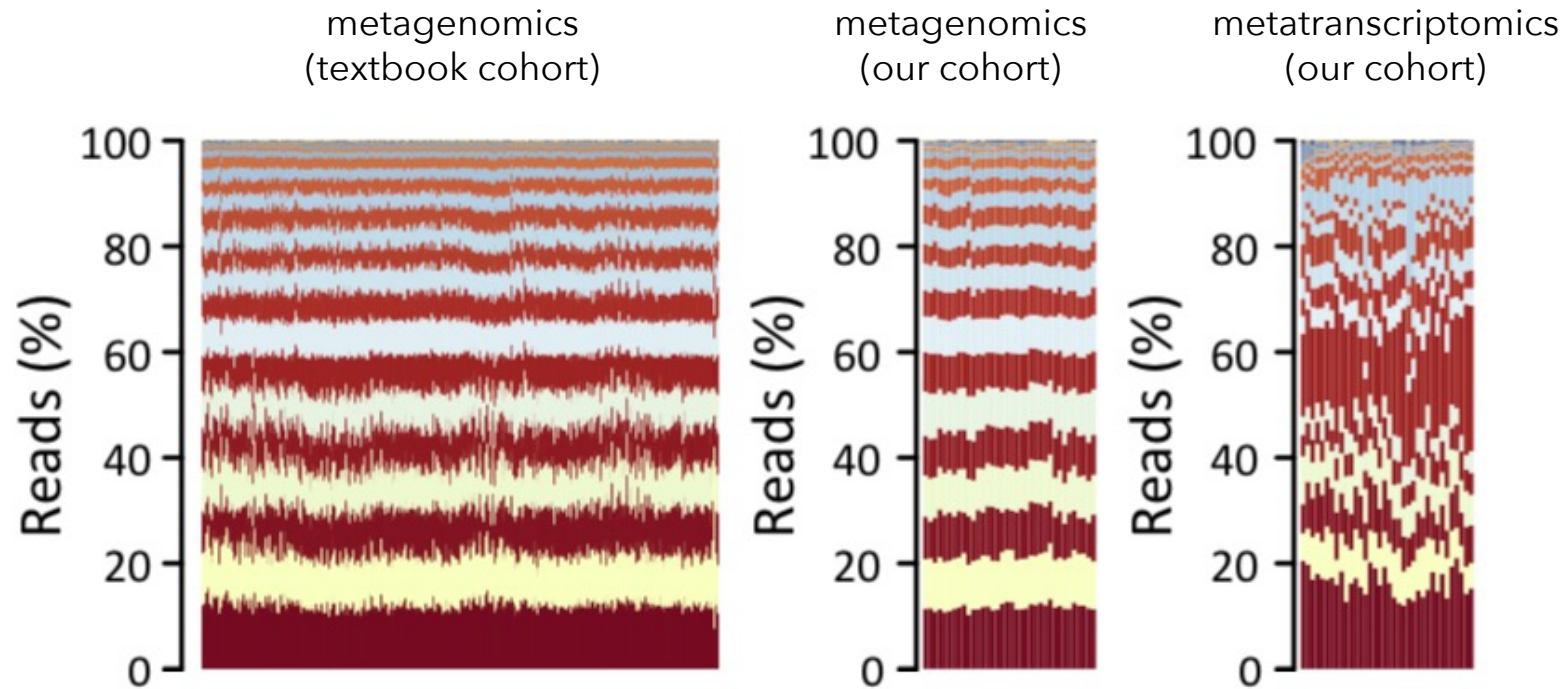


activity of populations within communities



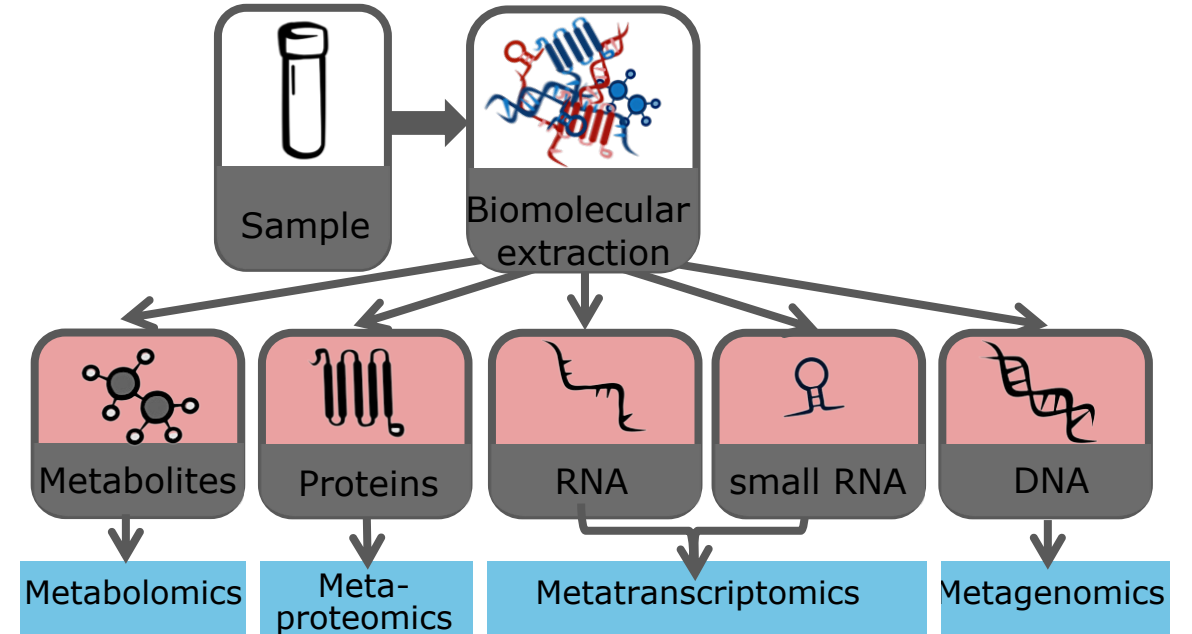
activity of community B

Metaomics

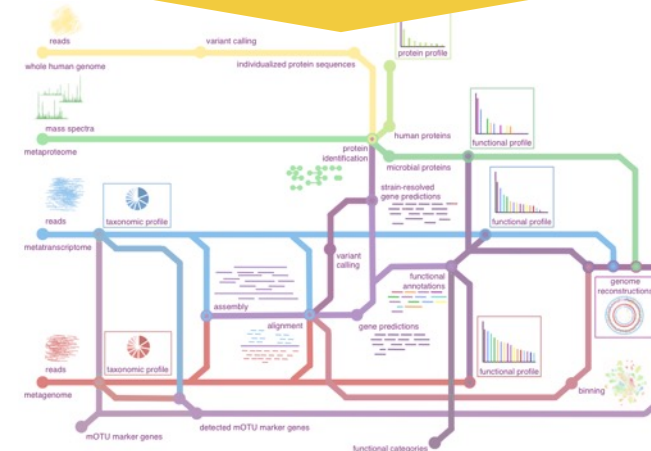


- metatranscriptomic profiles are much more variable

Sample to data



Data integration

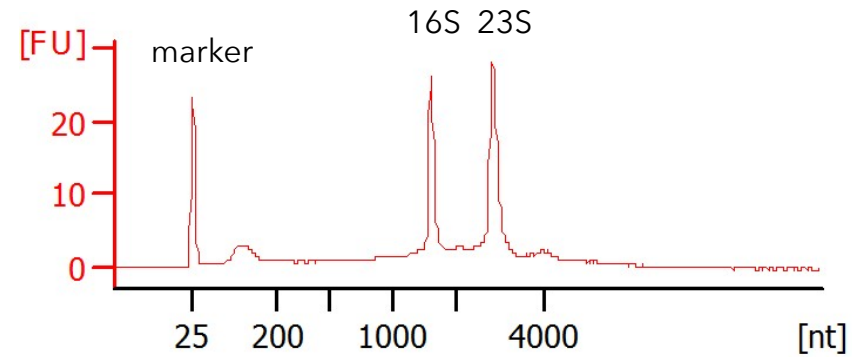
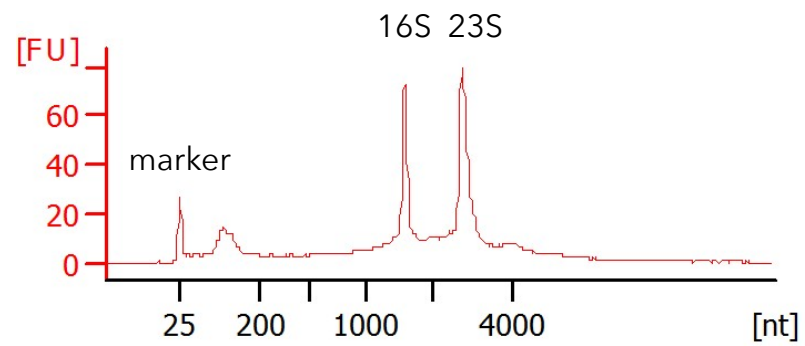
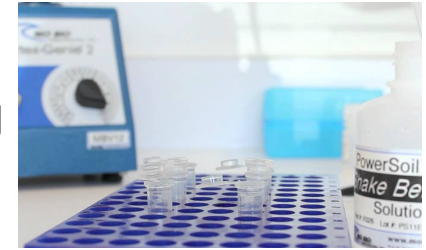


Sample to data

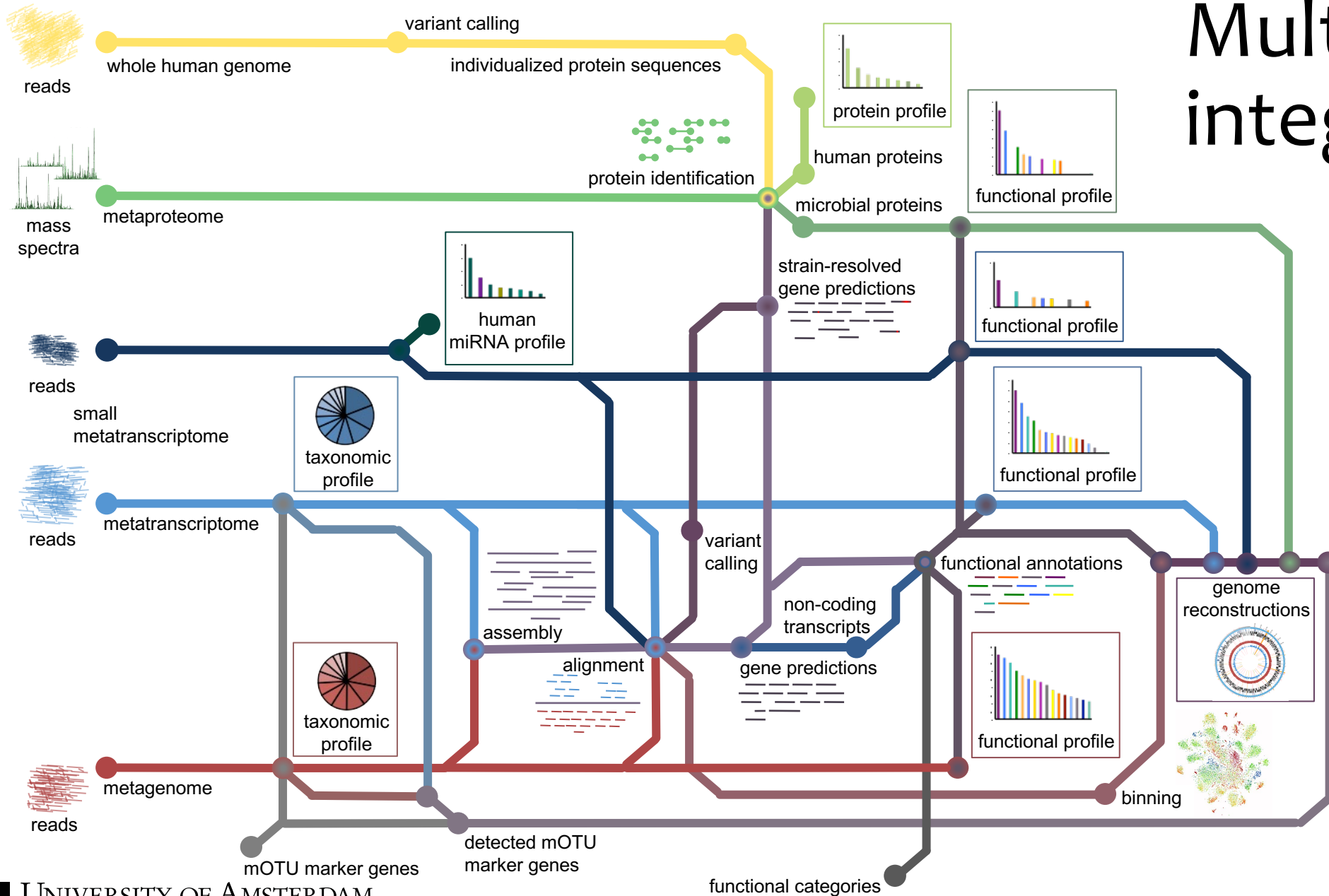
sampling + preservation



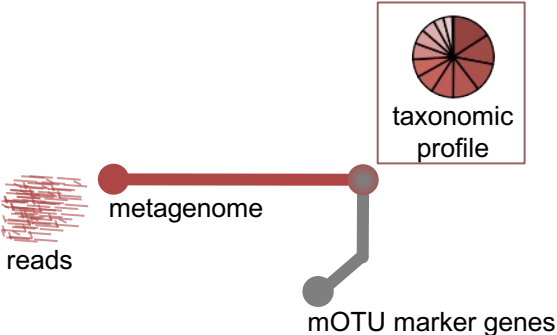
sample processing



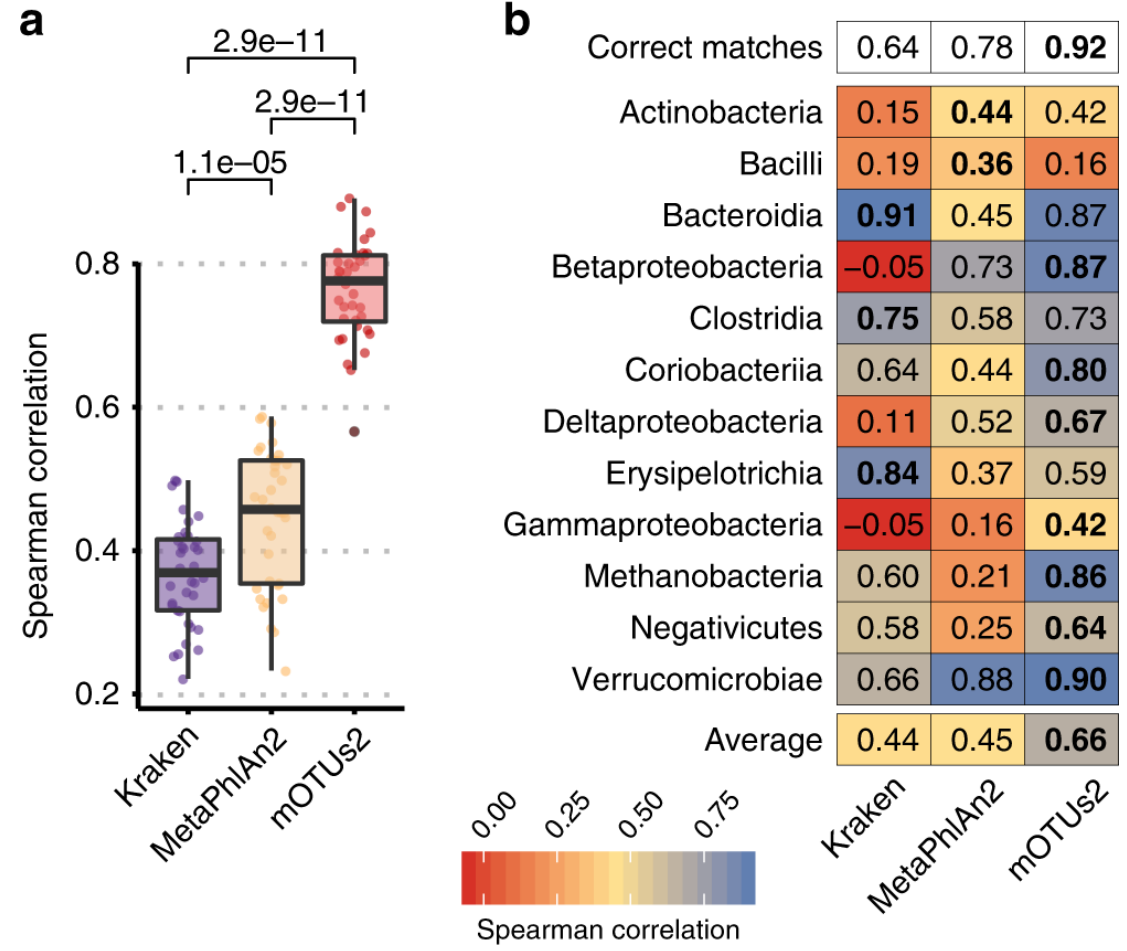
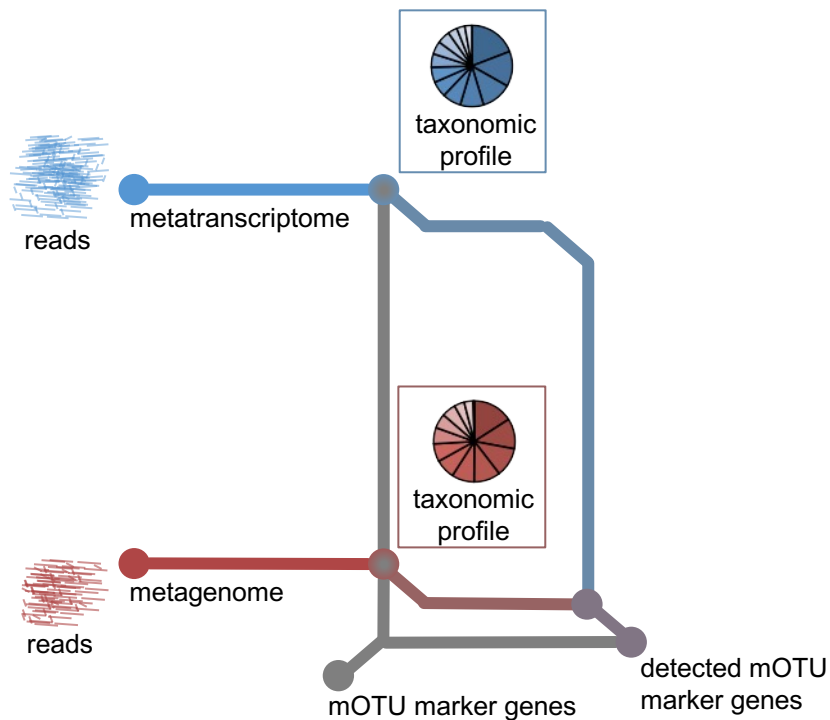
Multiomics integration



Workflow

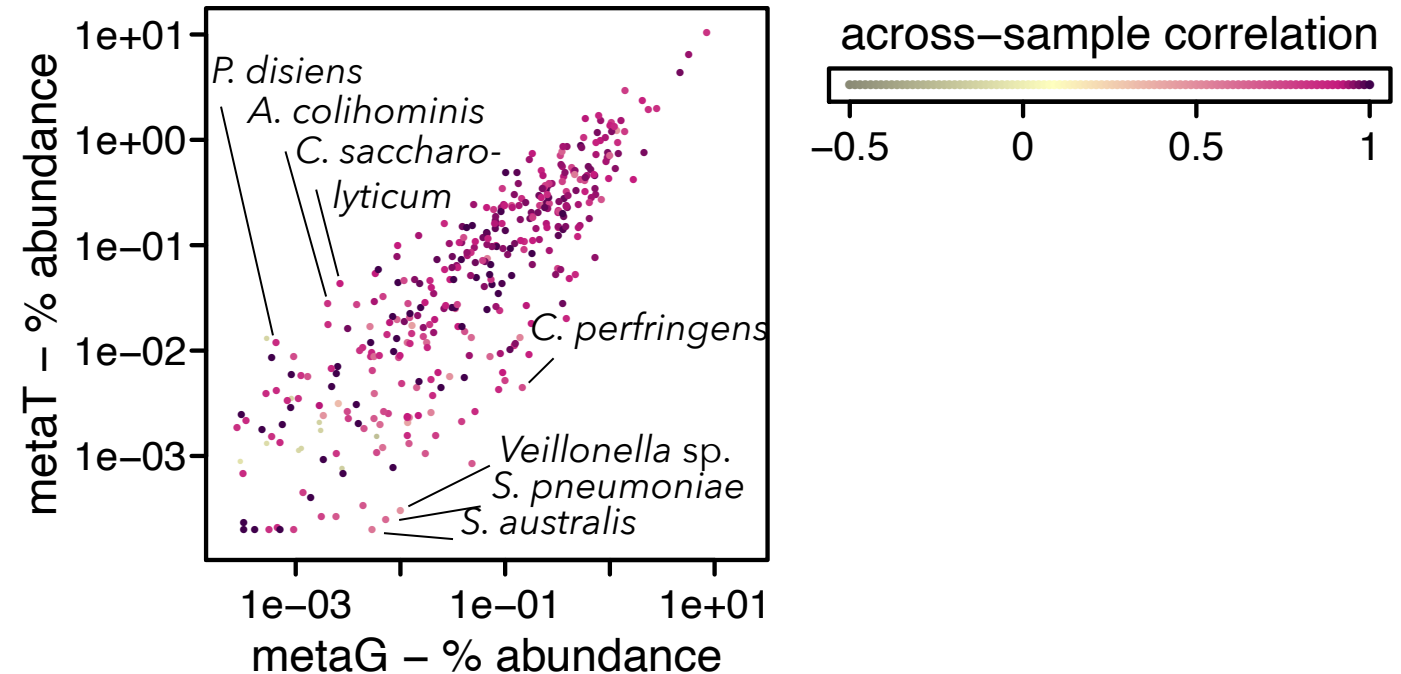
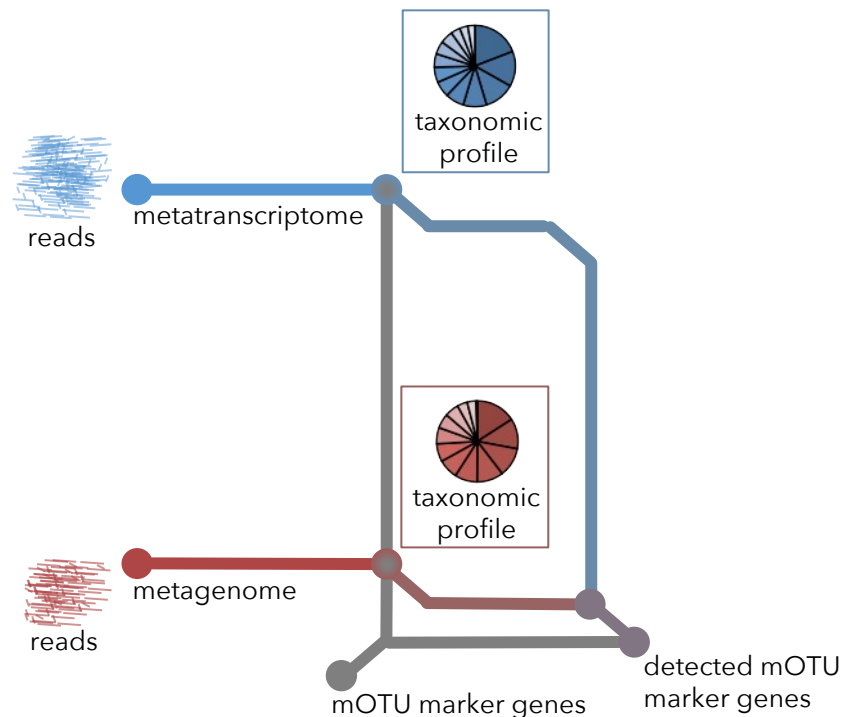
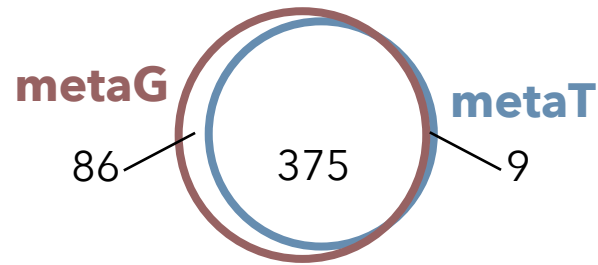


Taxonomic profiles?



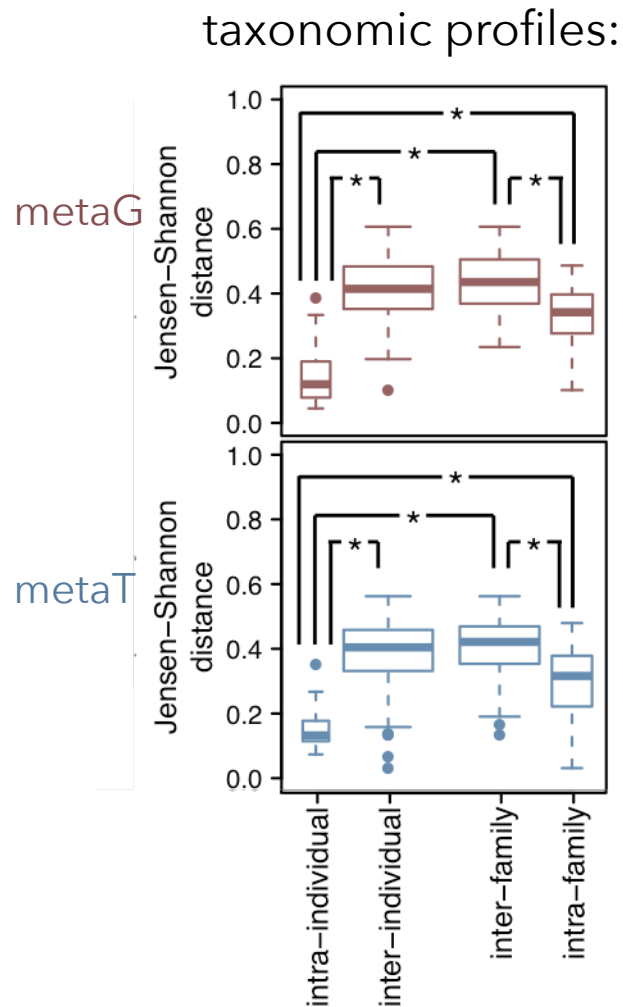
Milanese, A., Mende, D.R., Paoli, L. et al. Microbial abundance, activity and population genomic profiling with mOTUs2. Nat Commun 10, 1014 (2019). <https://doi.org/10.1038/s41467-019-08844-4>

MetaT vs metaG - taxonomic profiles

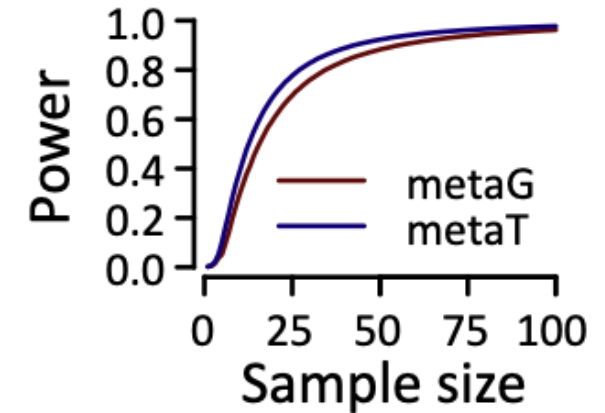
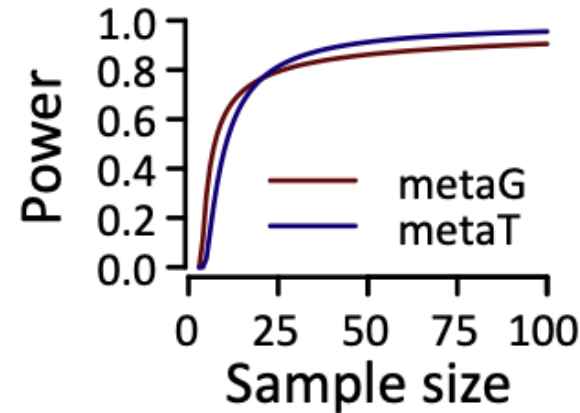


- **taxonomic profiles** at metagenomic and metatranscriptomic levels are **very similar**

Patterns

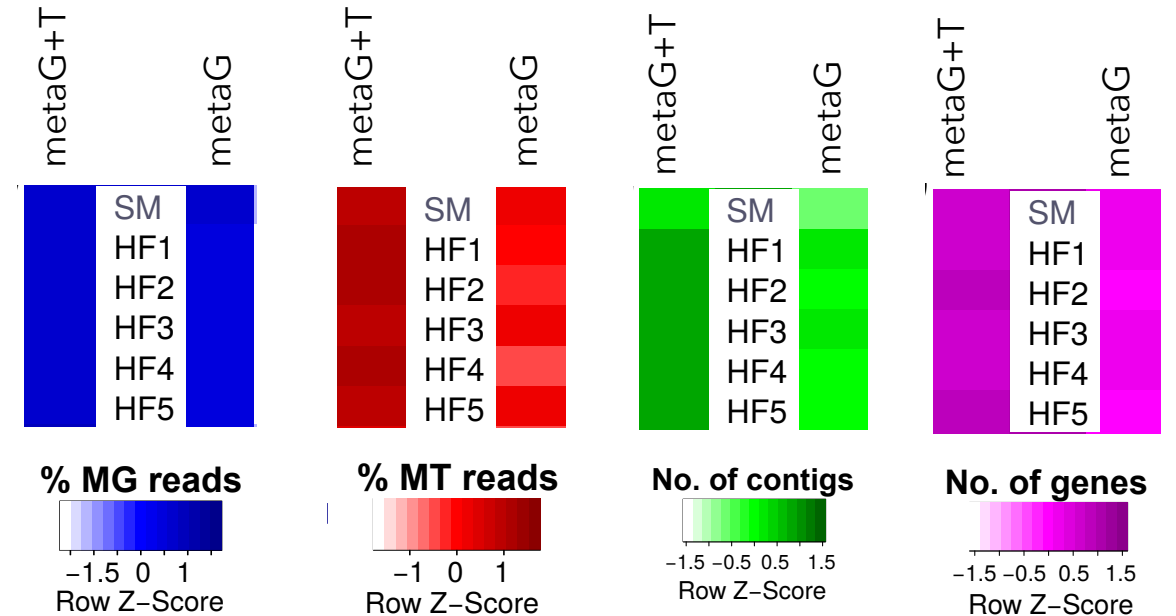
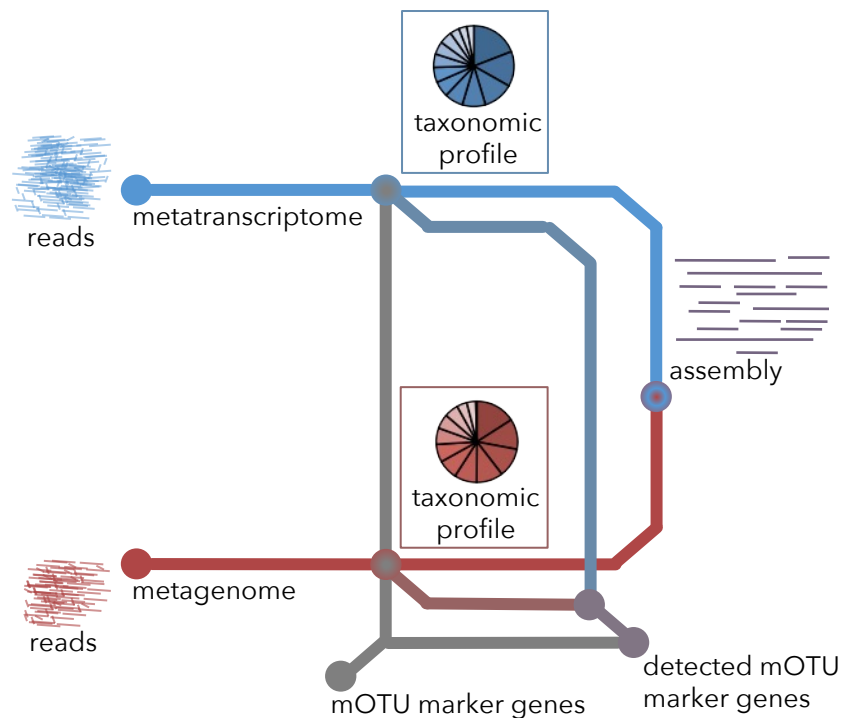


- metatranscriptomics-based taxonomic profiles show similar results to metagenomics



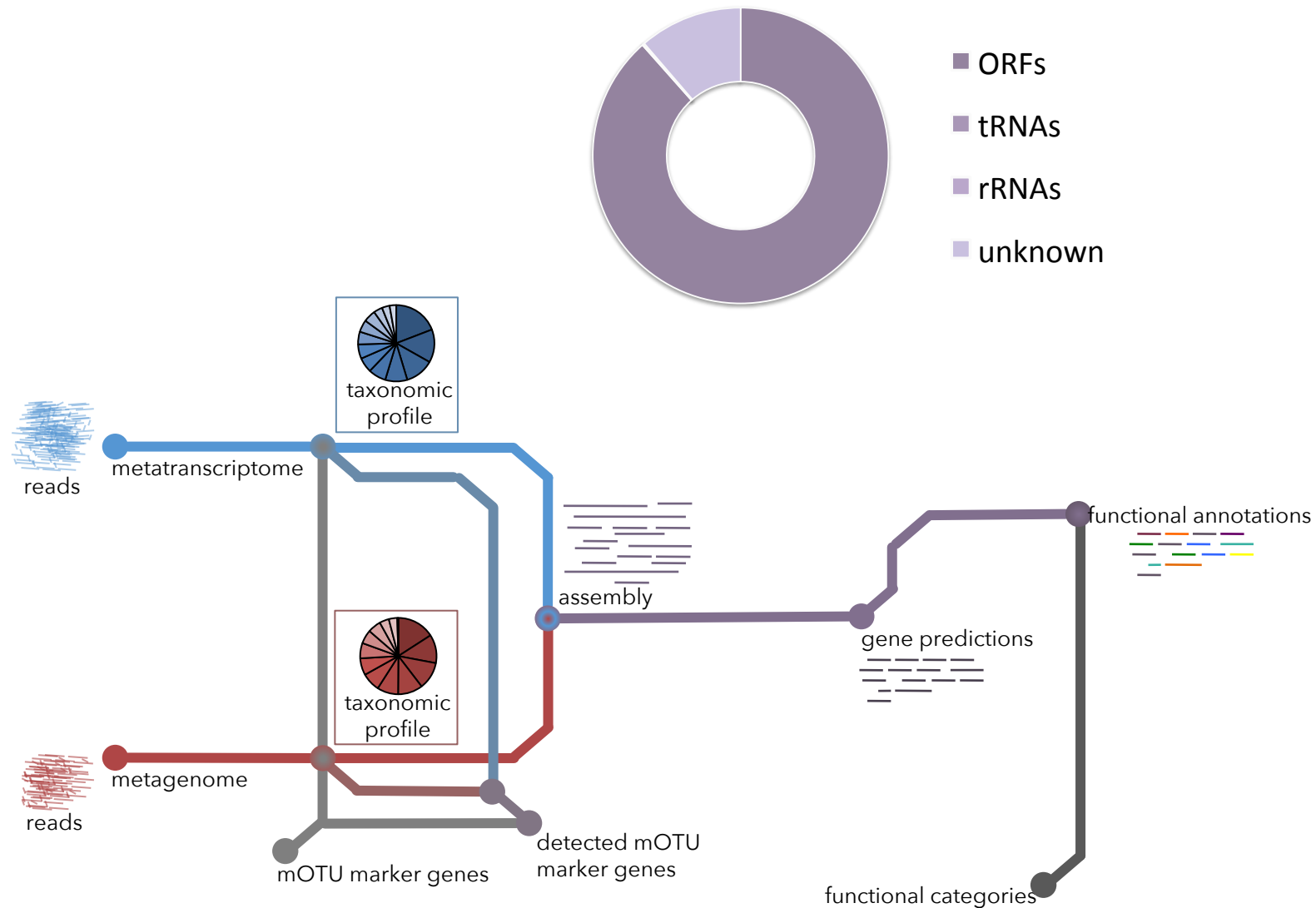
- metatranscriptomics have similar power to metagenomics

Workflow

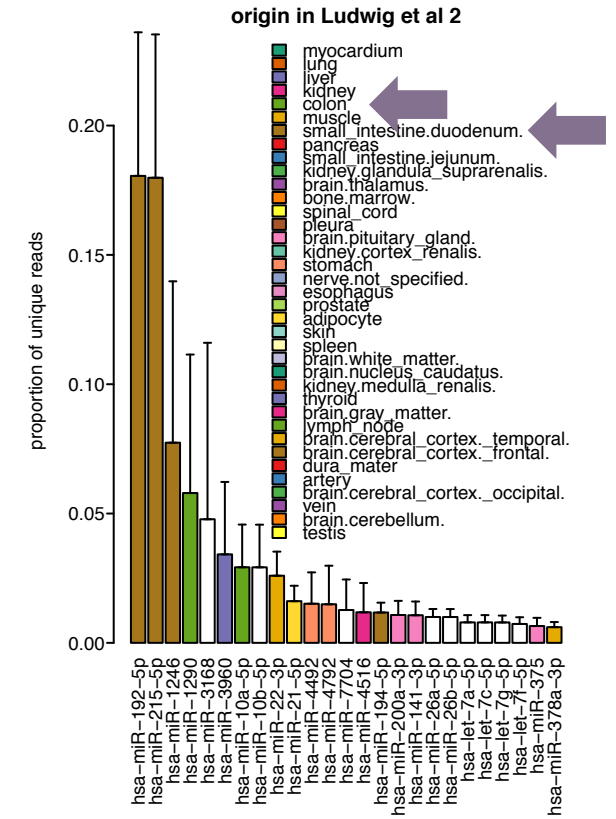
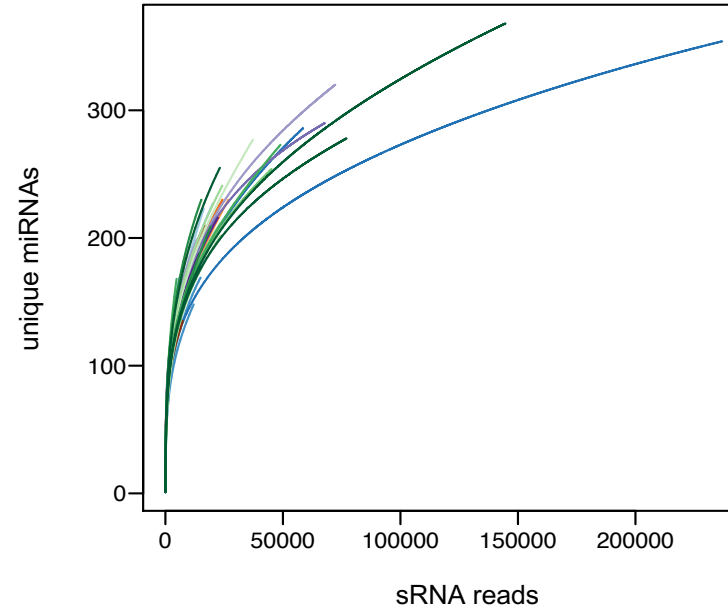
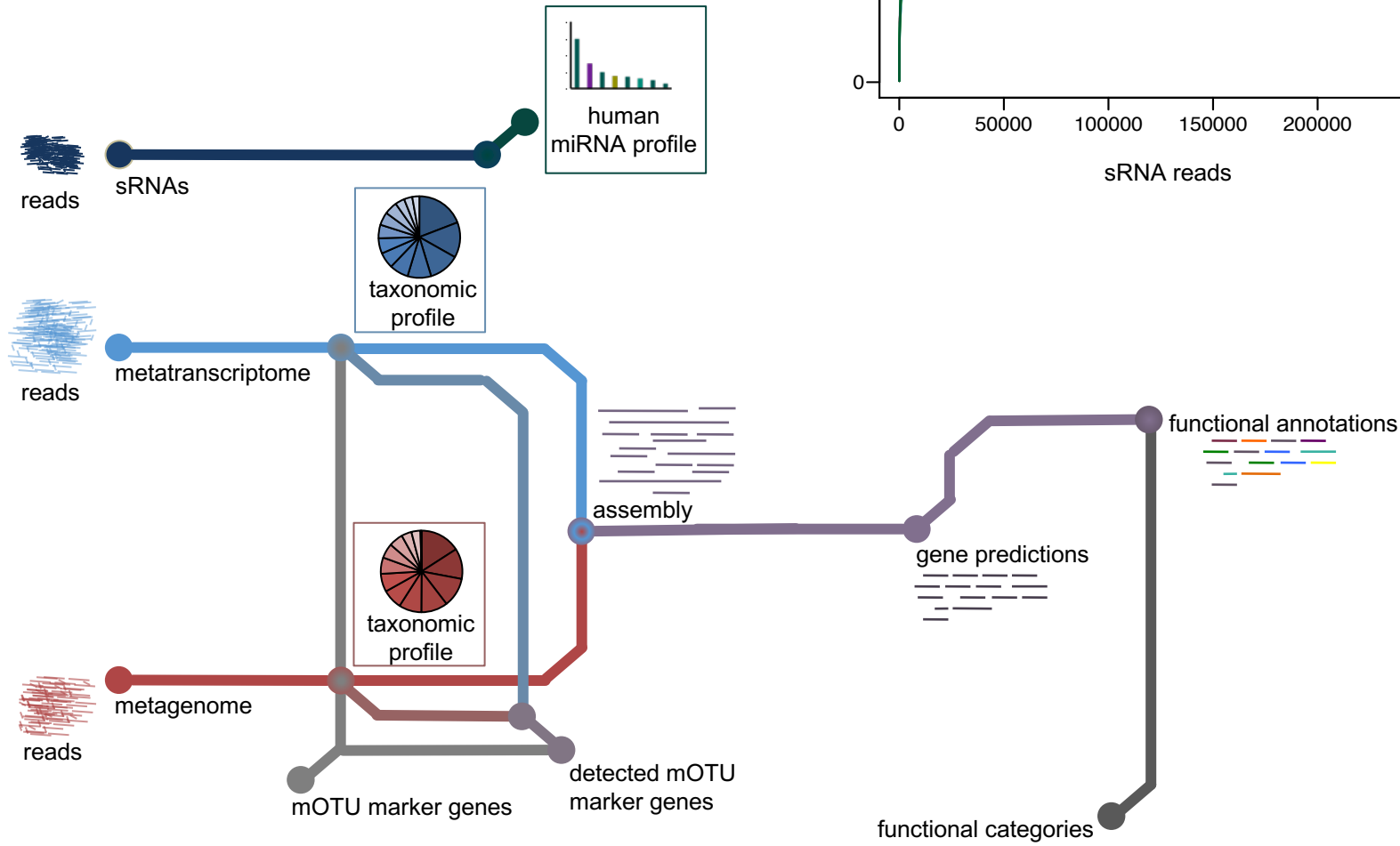


- improved data usage
- better contiguity
- more ORFs

Workflow

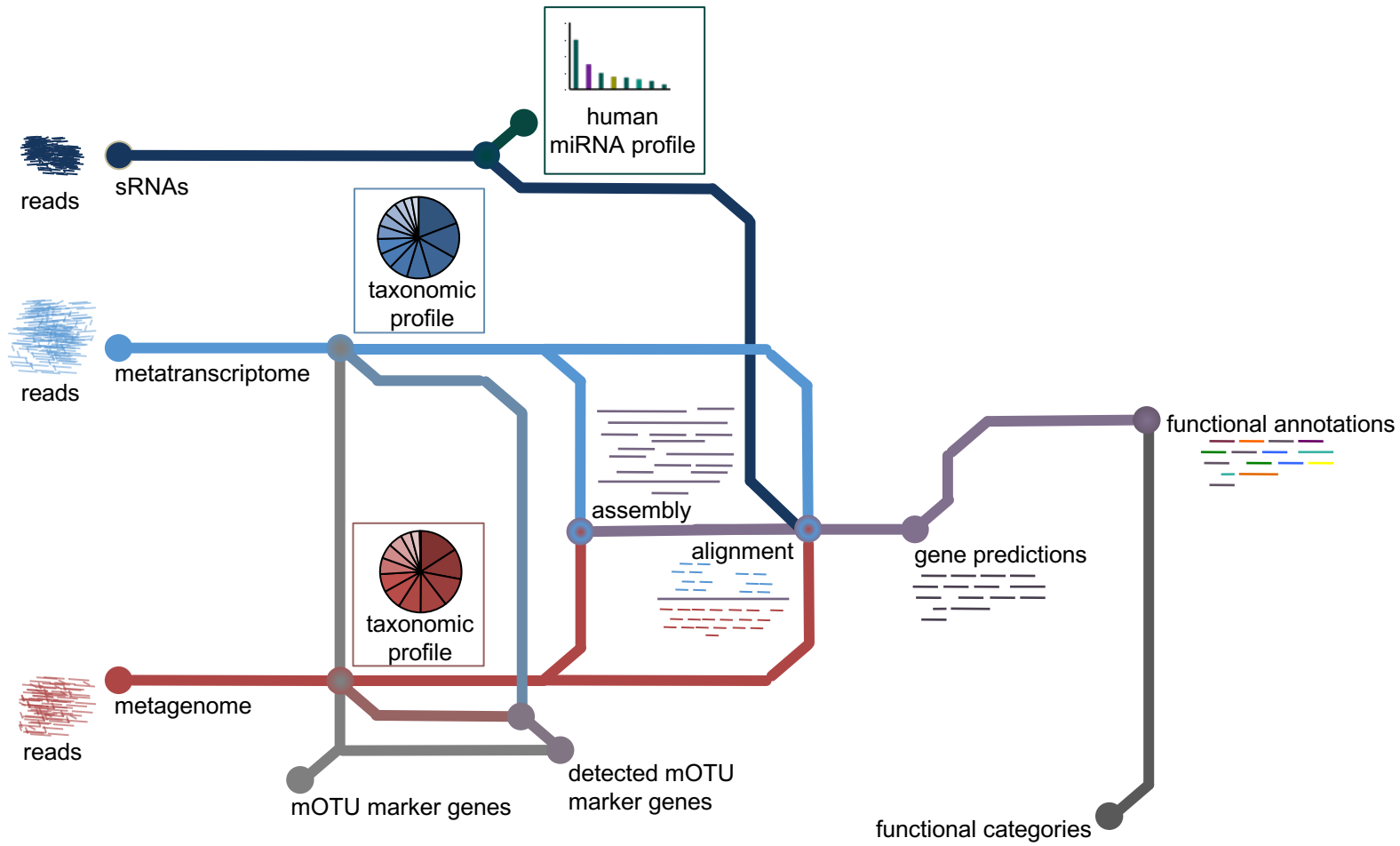


Workflow

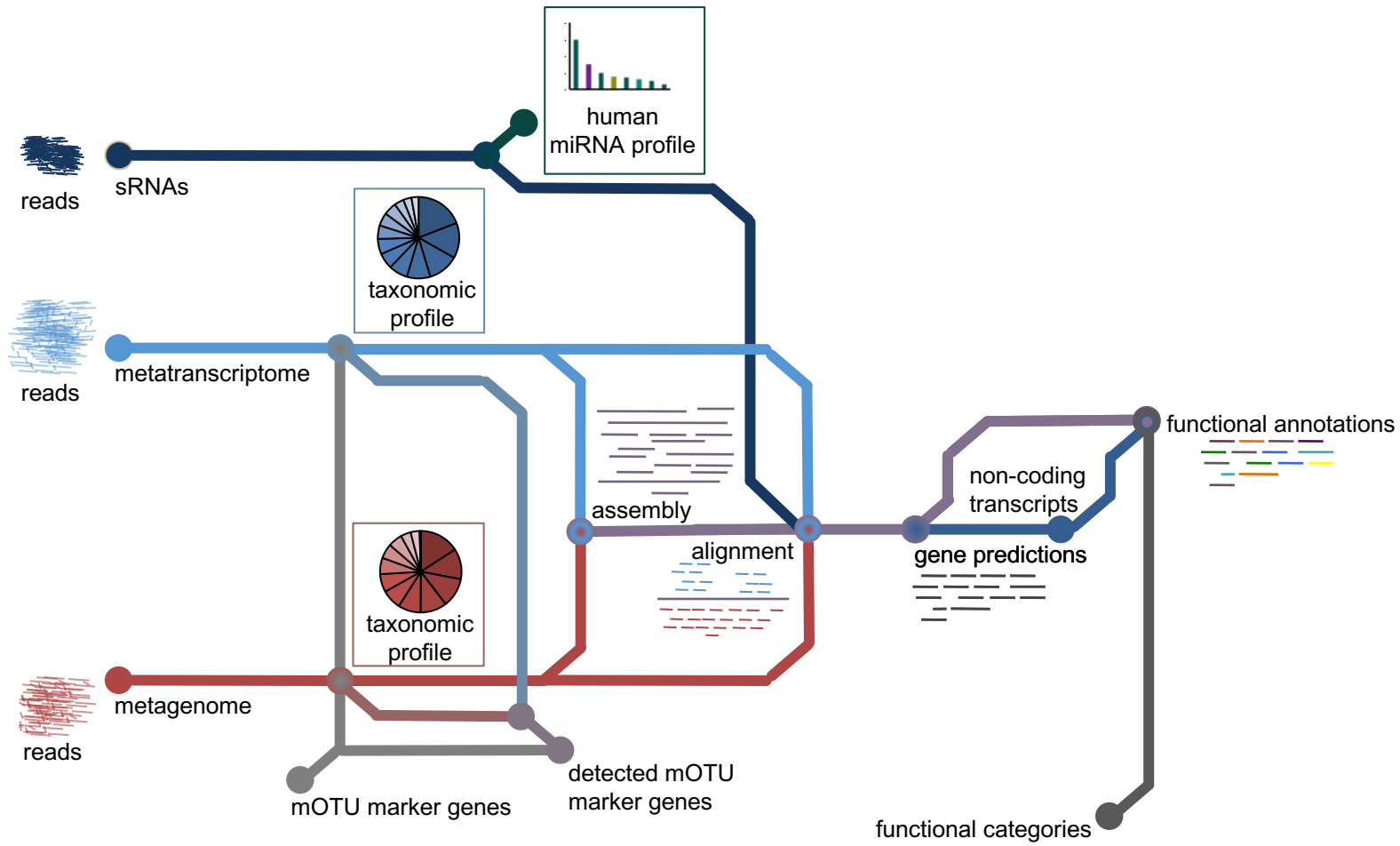


Ludwig *et al.*, Distribution of miRNA expression across human tissues. *Nucleic Acids Res.* 2016 May 5;44(8):3865-77. doi: 10.1093/nar/gkw116

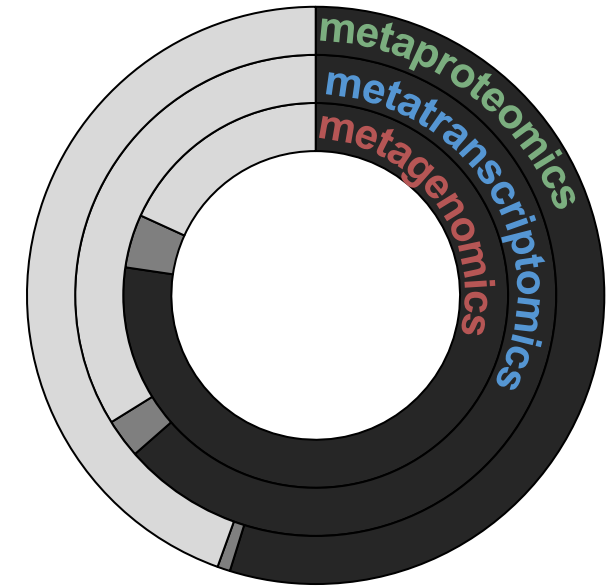
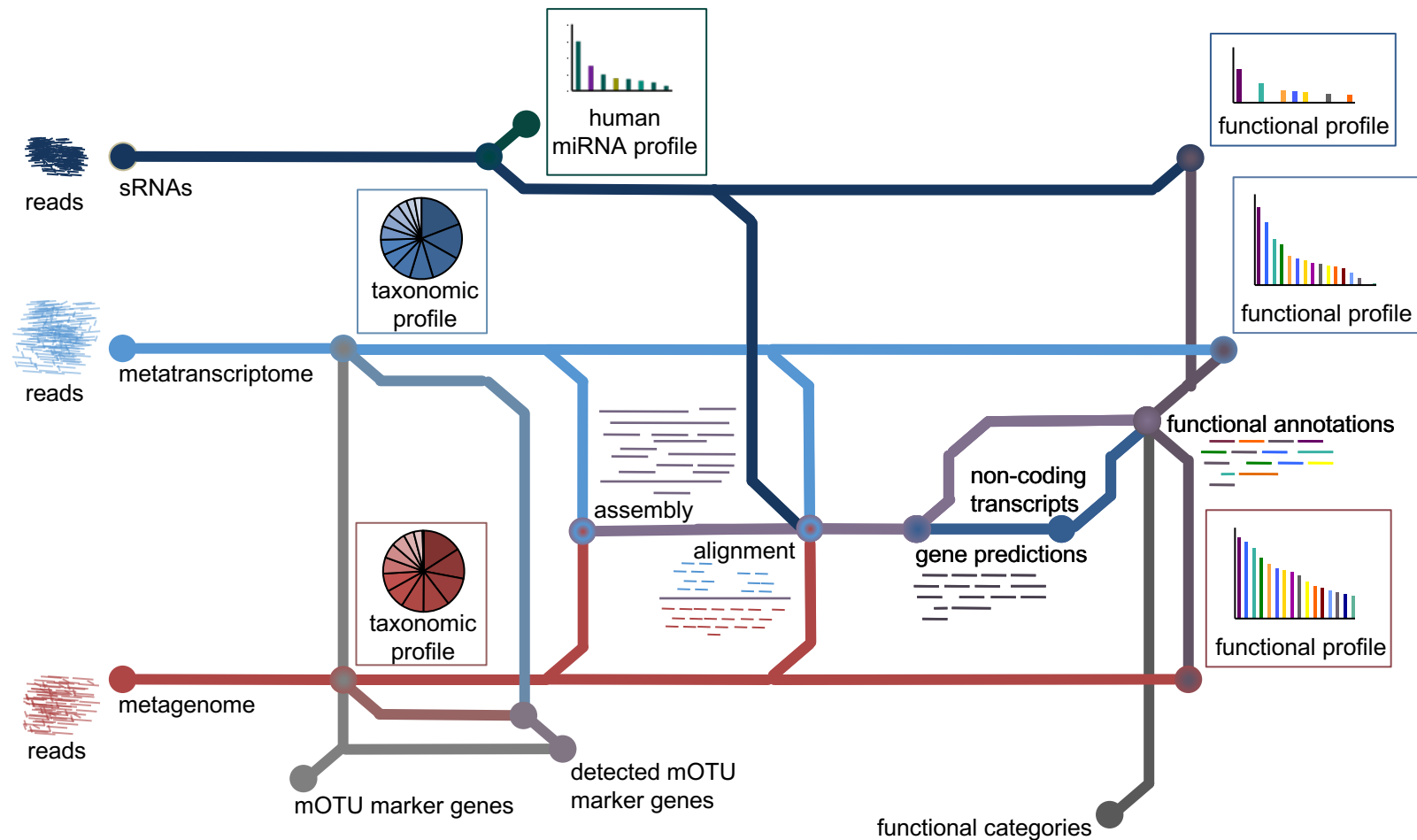
Workflow



Workflow

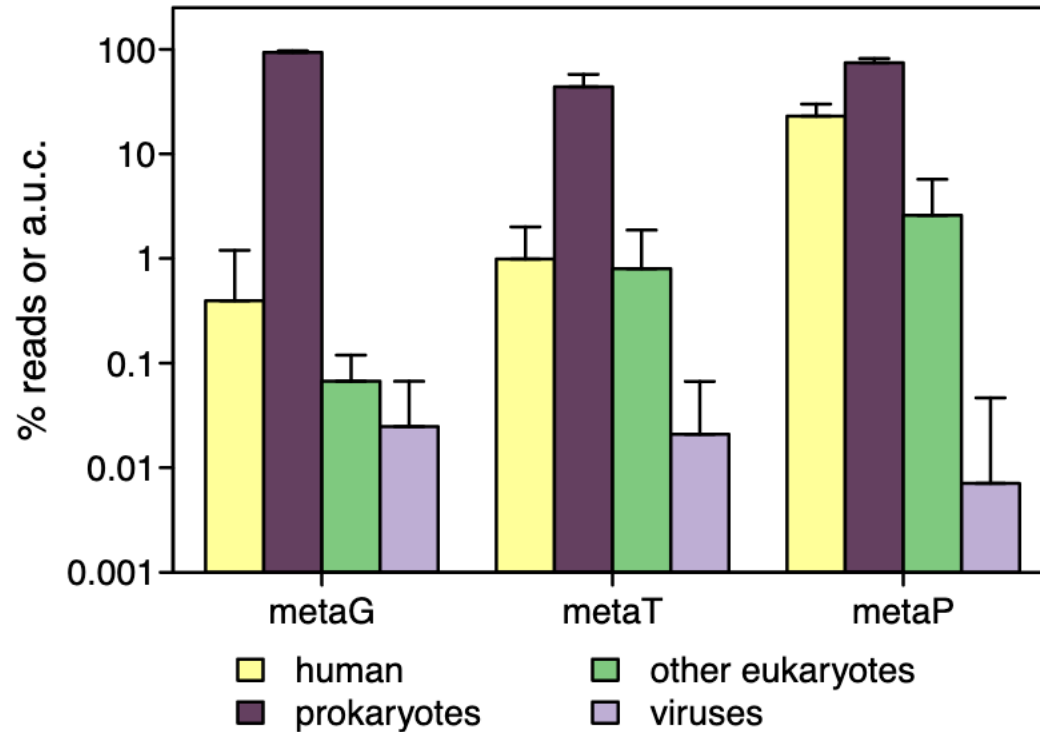


Workflow



- function predicted
- ▒ domains of unknown function
- no function predicted or uncertain identification

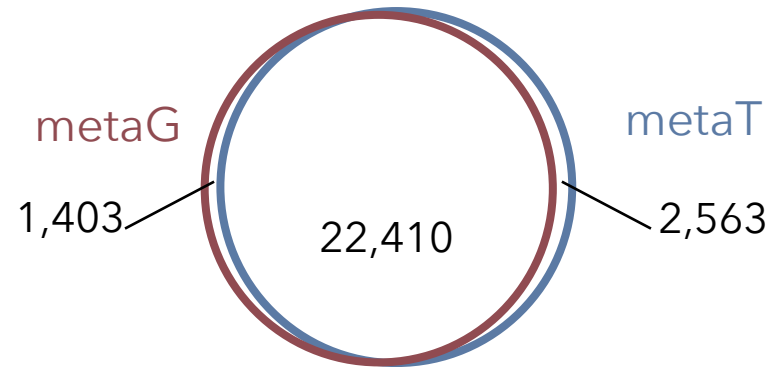
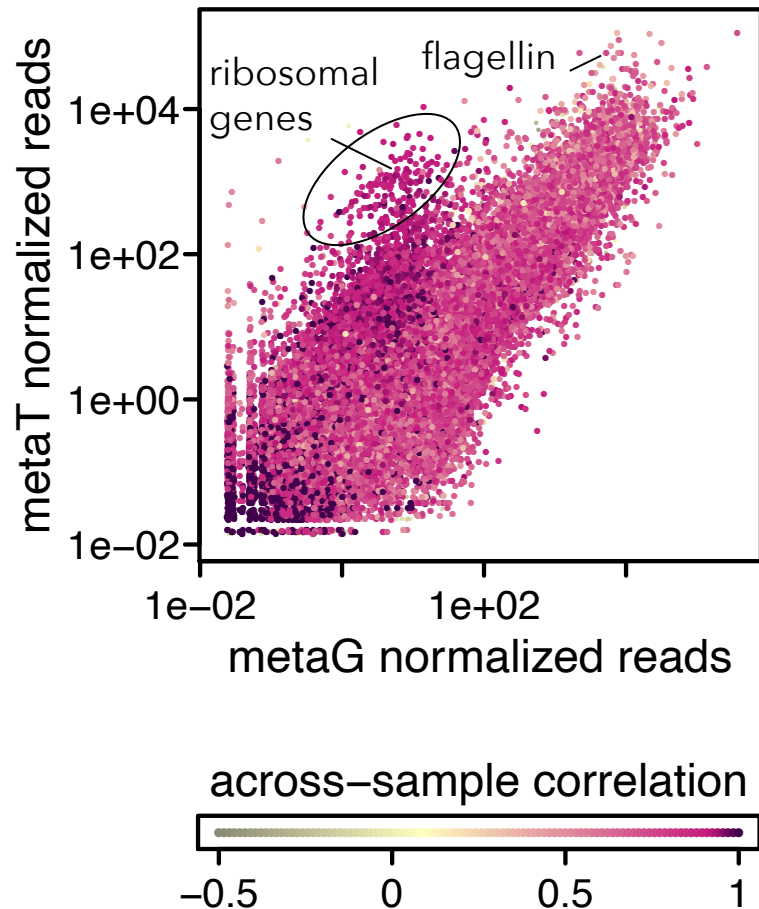
MetaT vs metaG - what do we detect?



- overall similar origins, but larger organisms are more represented
- often more host!

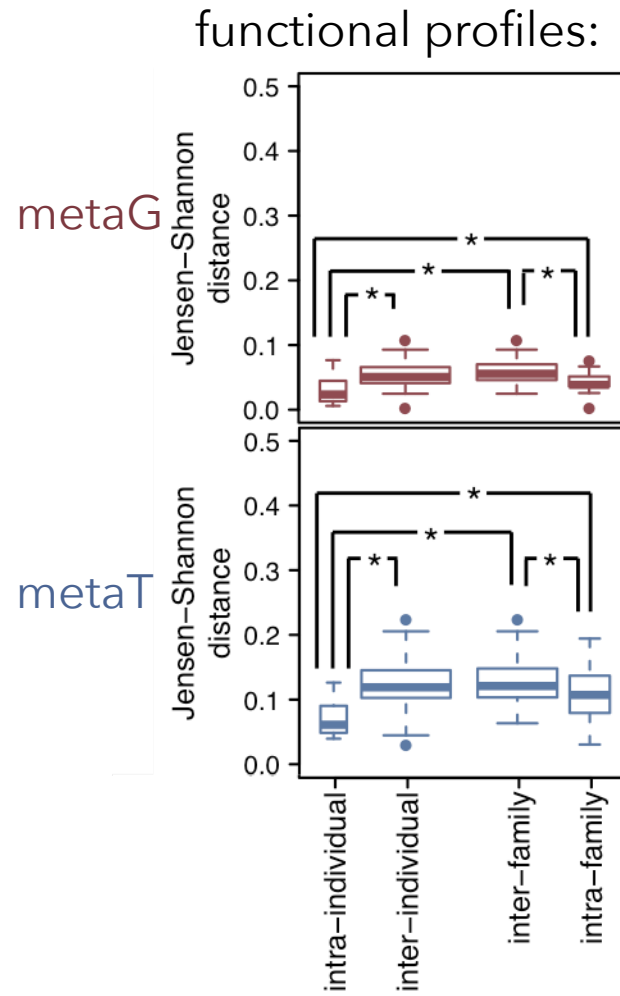
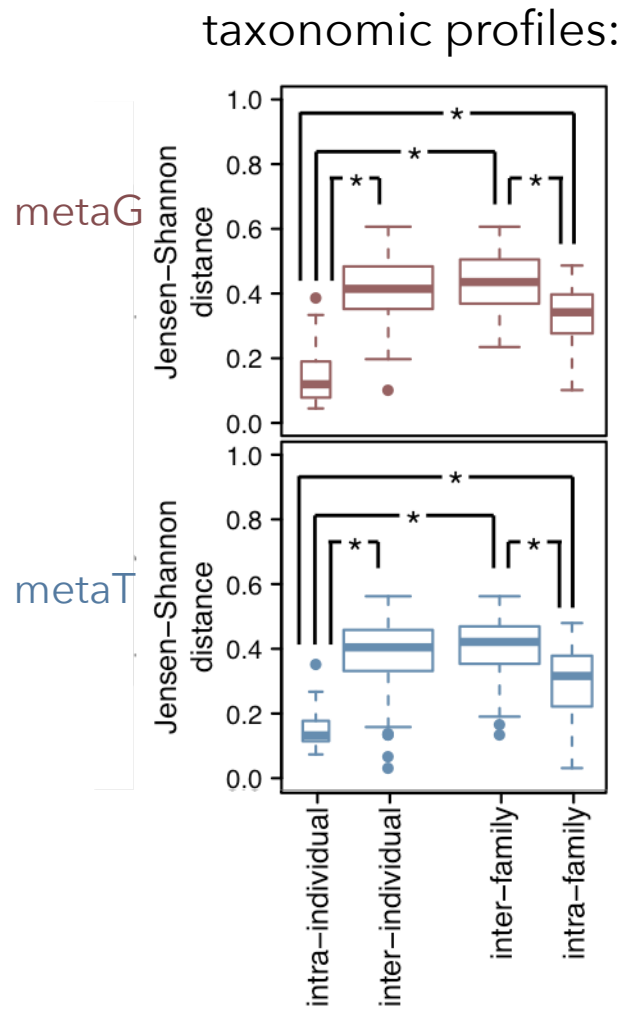
MetaT vs metaG - functional profiles

functional profiles:

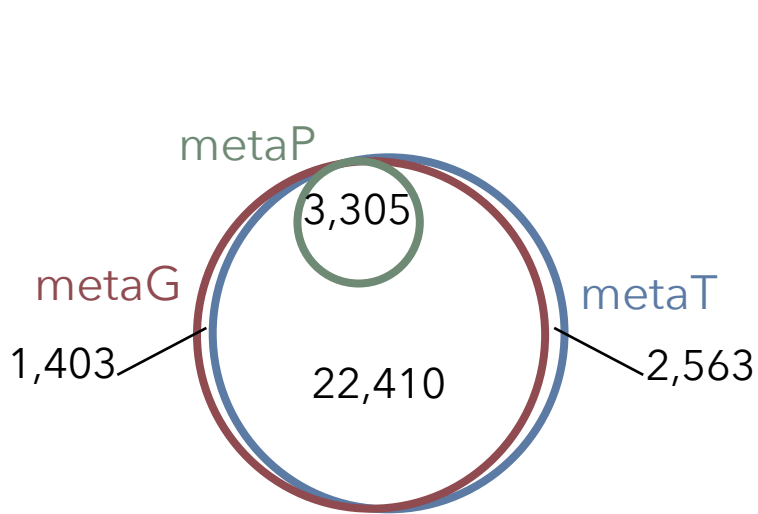


- **functional profiles** at metagenomic and metatranscriptomic levels are **less similar than taxonomic profiles**

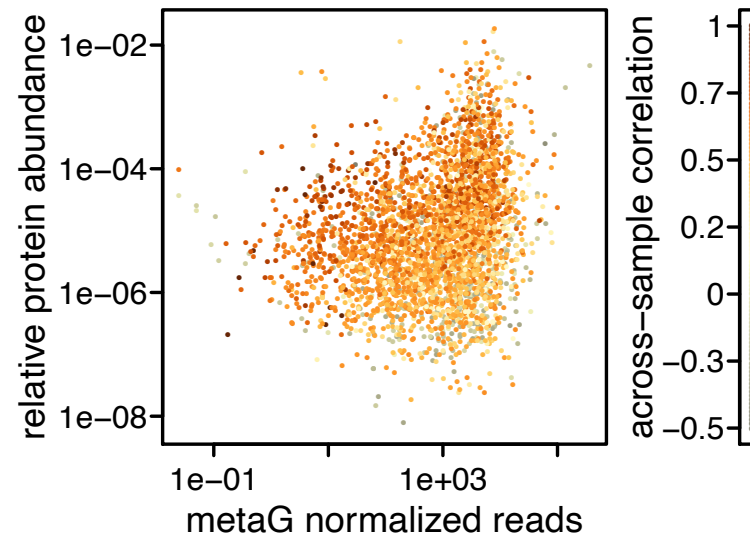
Patterns



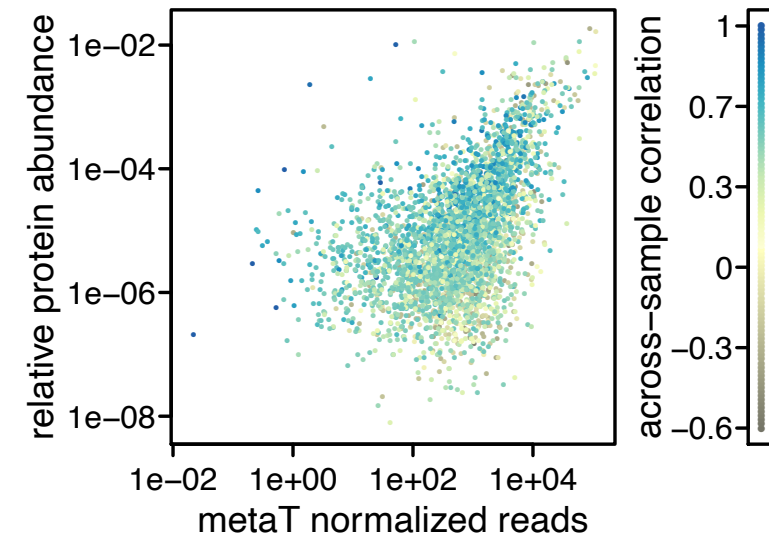
MetaT vs metaP - functional profiles



metaP vs metaG:

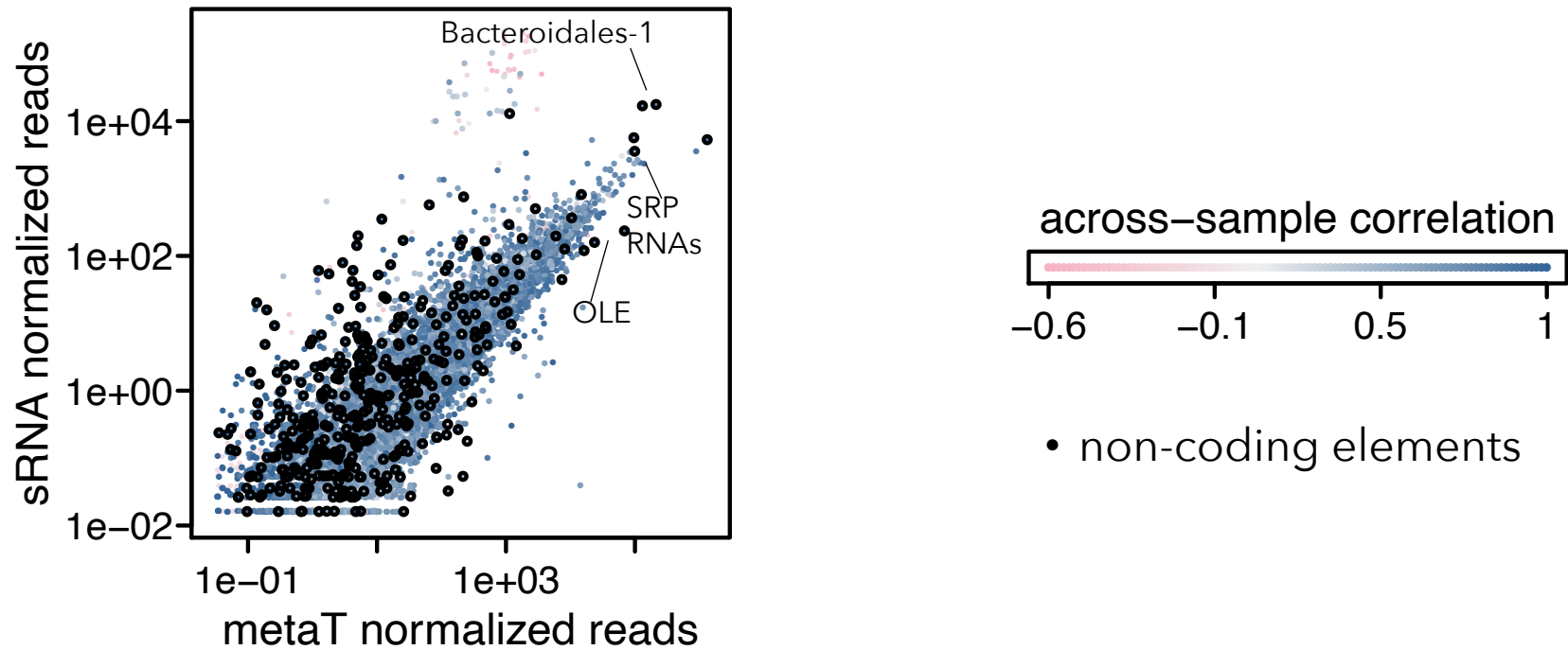


metaP vs metaT:



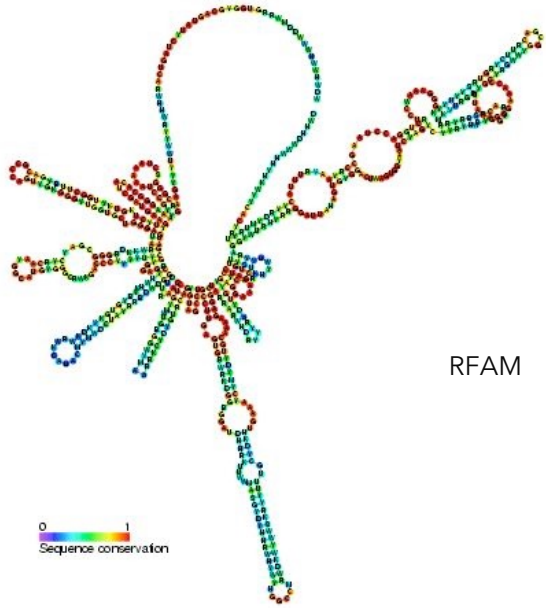
- metaproteome is more related to metatranscriptome

Coding and non-coding transcripts



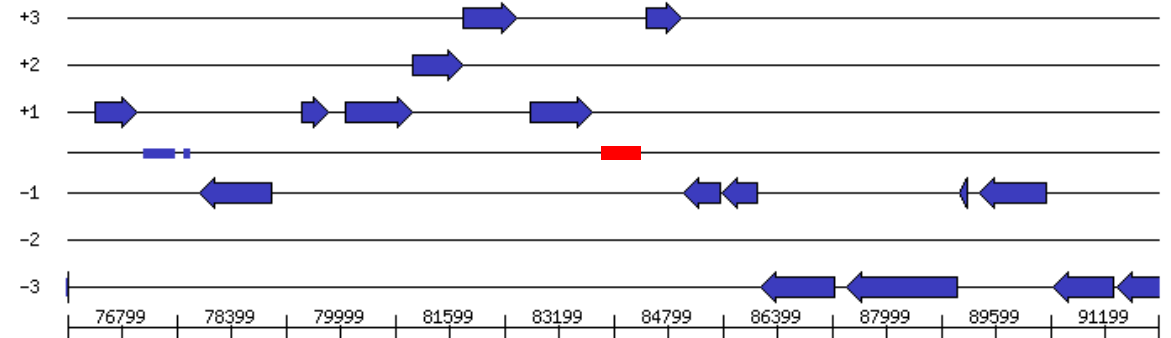
- some highly expressed transcripts are non-coding

Example - OLE (ornate, large, extremophilic) RNA

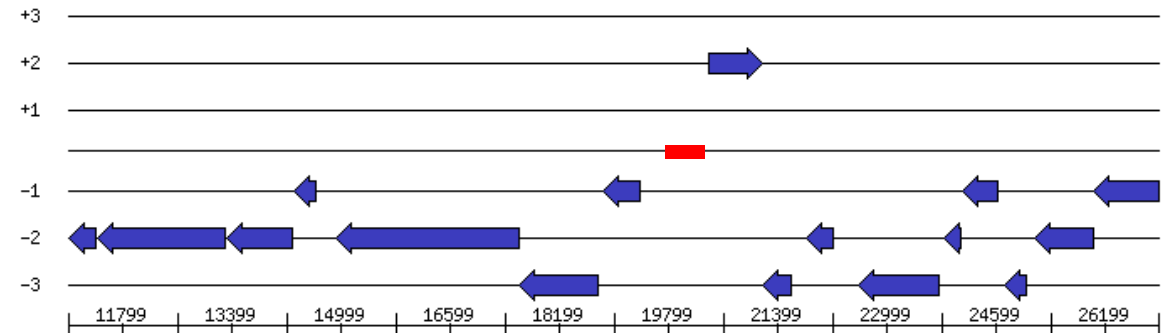


- found in Firmicutes
- ~ 600 nt length

Firmicutes mOTU:

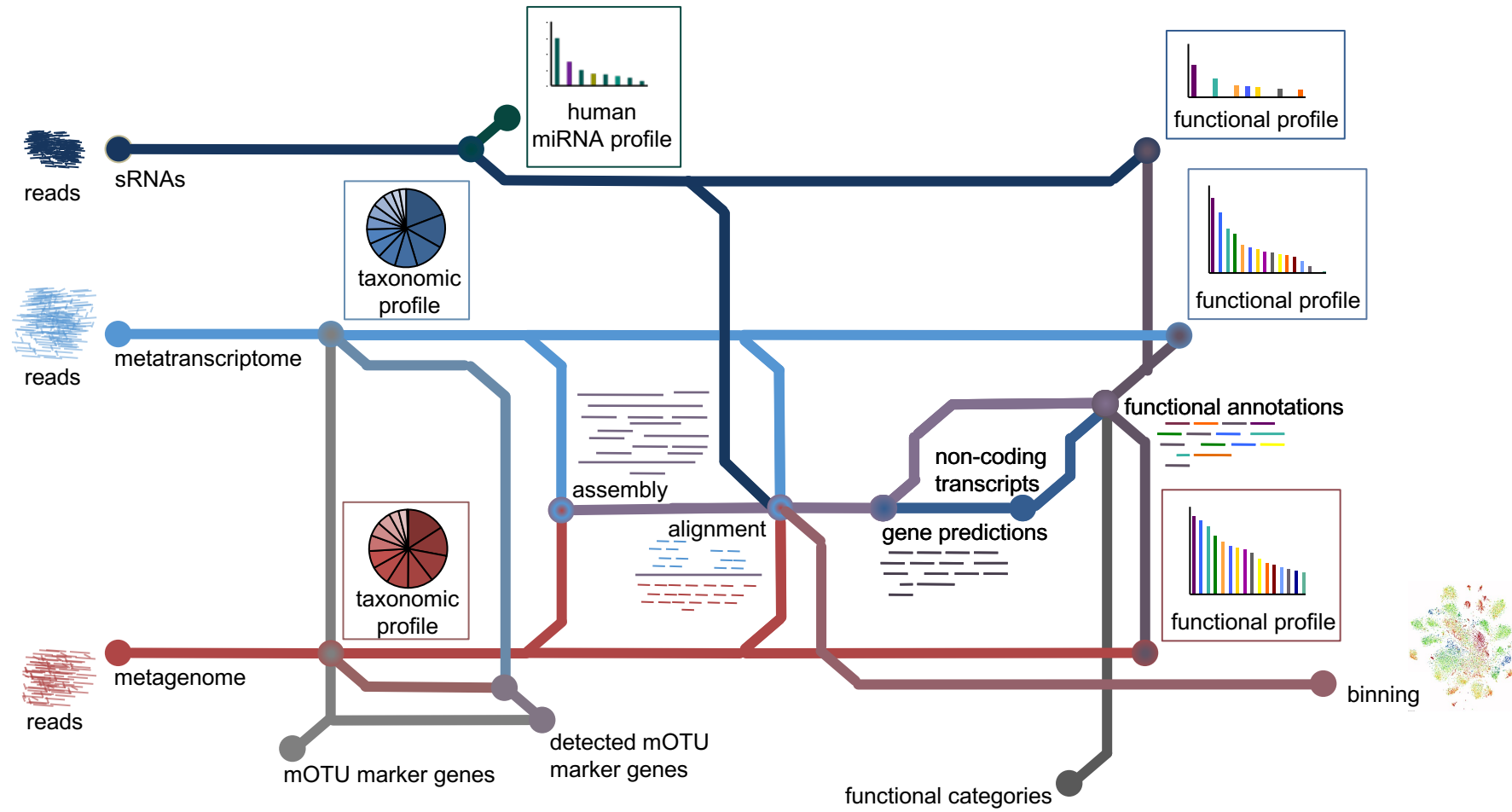


Clostridia mOTU:

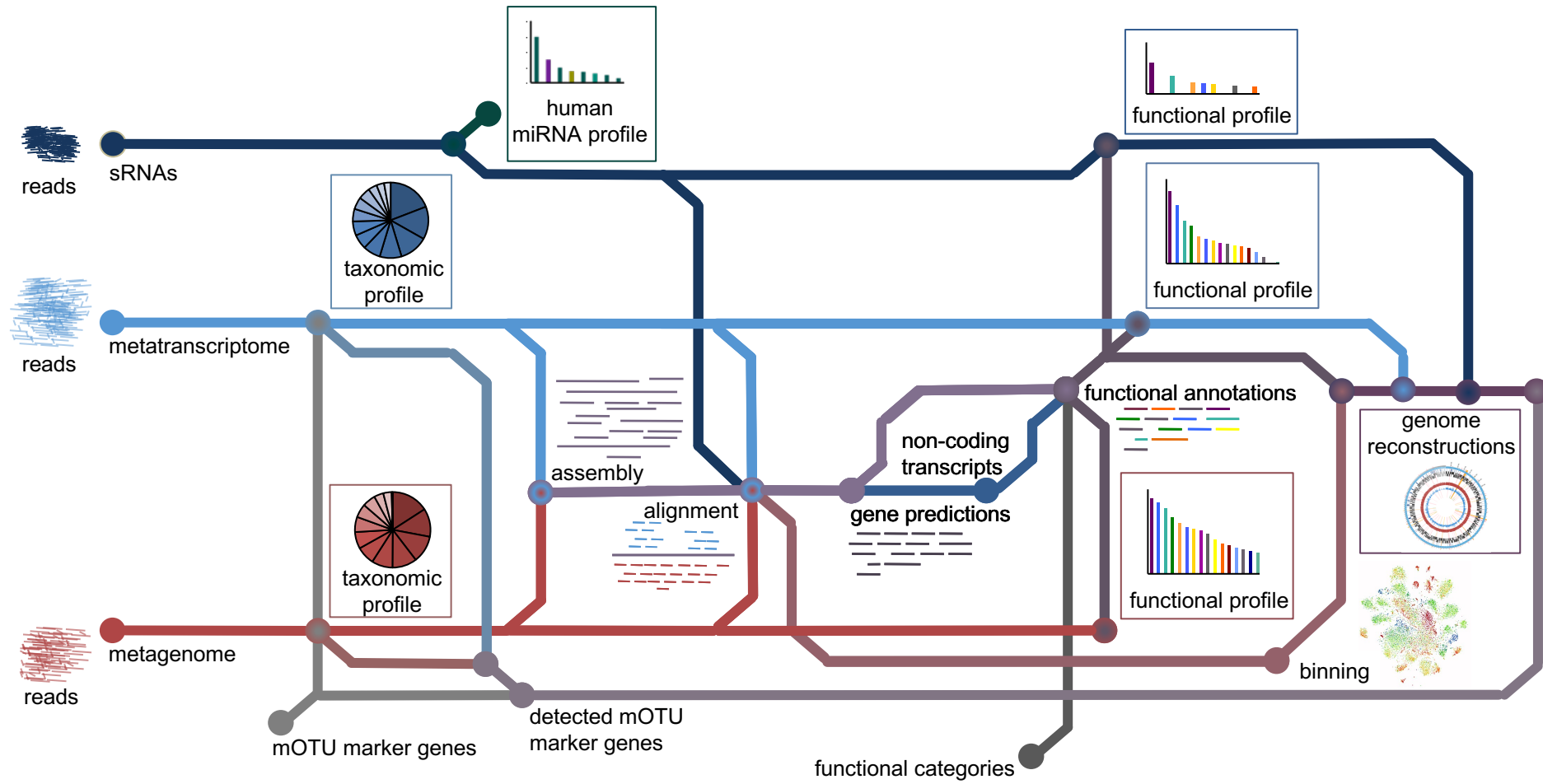


Heintz-Buschart *et al.*, unpublished

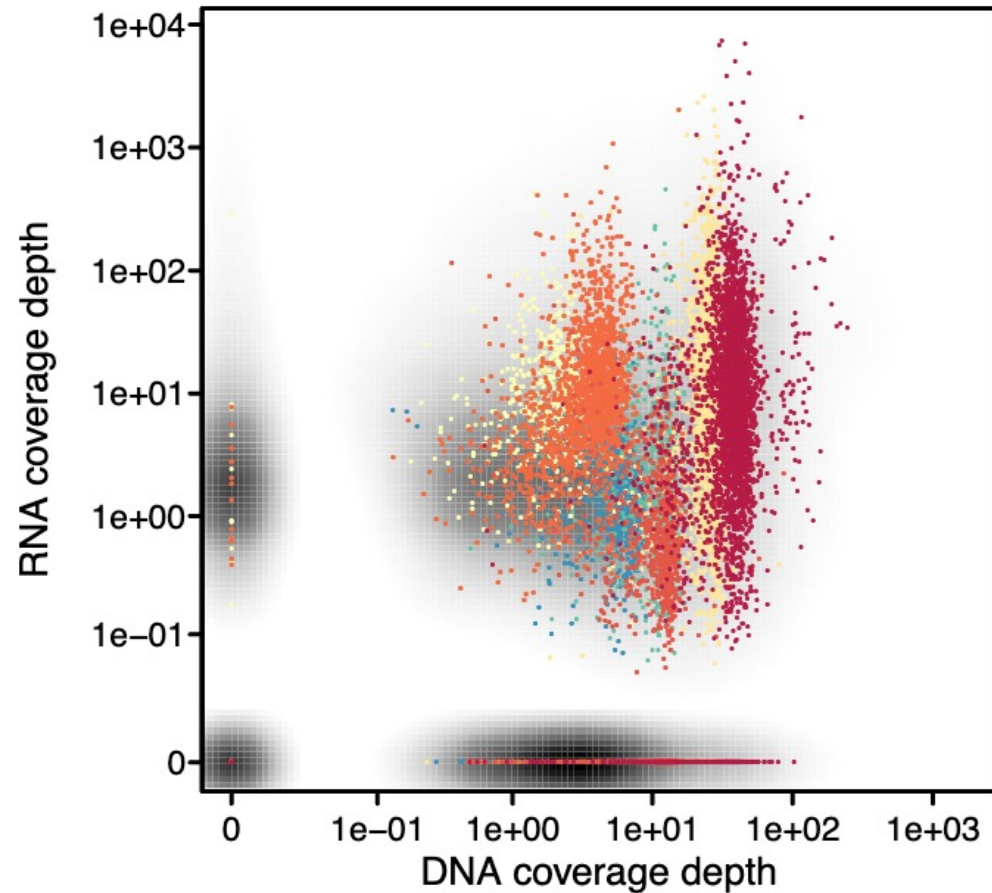
Workflow



Workflow



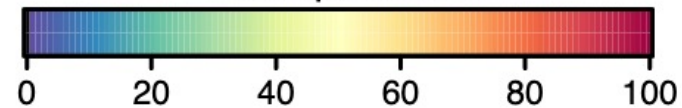
Expression per genome



binned population-level genomes – taxonomy

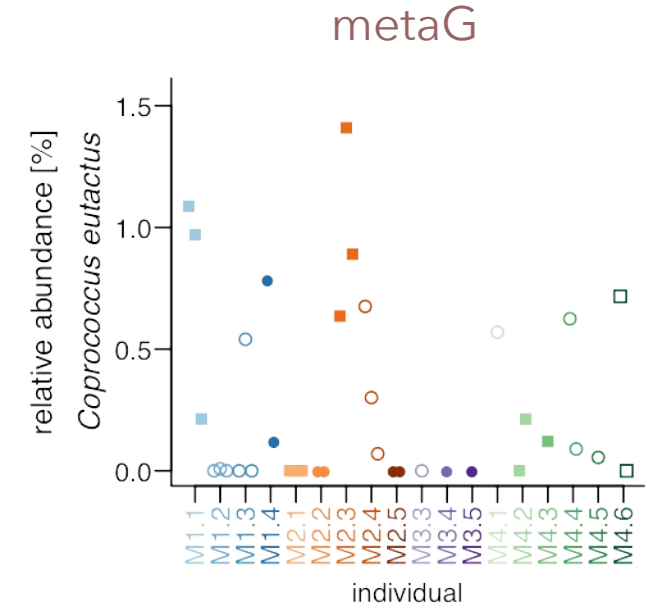
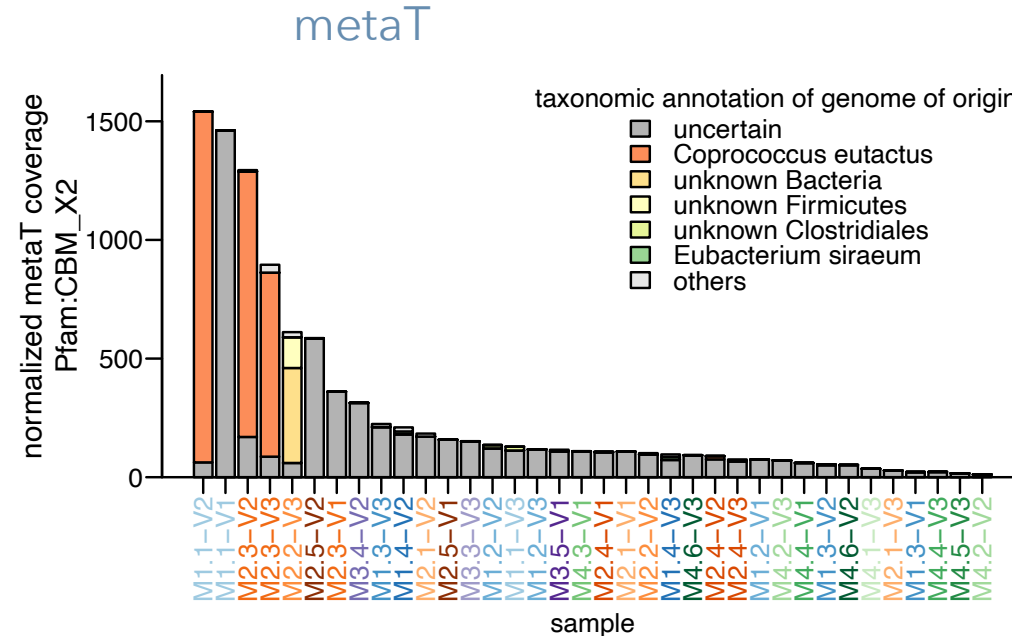
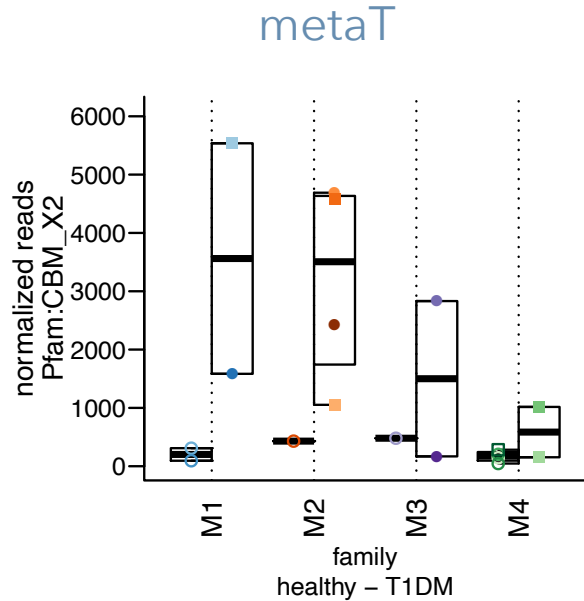
- G2.2.2 – *Butyrivibrio crossotus*
- G11 – Firmicutes
- G31 – unclassified Clostridiales
- O32 – unclassified Clostridiales
- O50 – *Ruminococcus* sp.
- C1.1.2 – *Collinsella aerofaciens*
- C24.1 – unclassified Clostridiales

% completeness



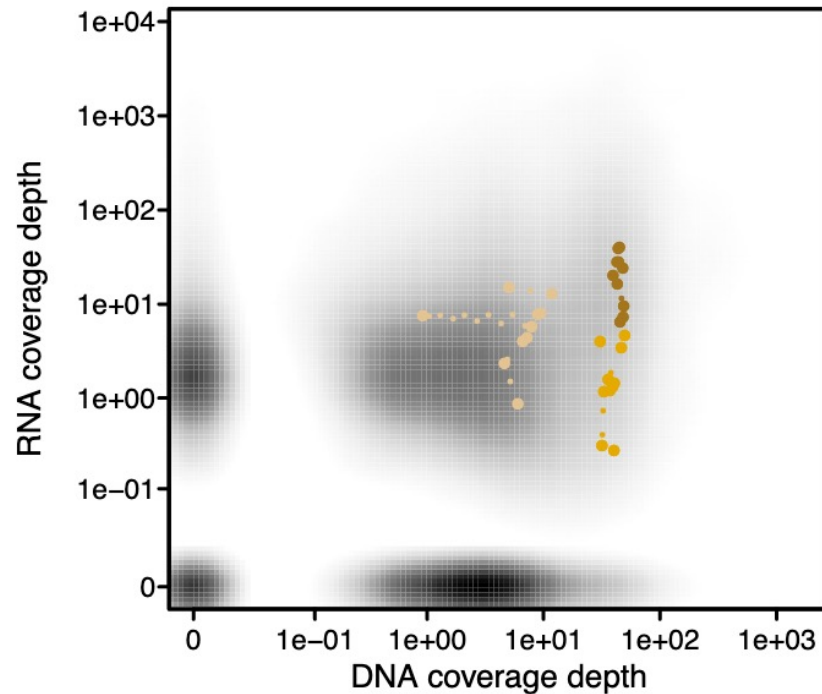
Example - who expresses a gene of interest?

example: protein with cellulose-binding domain



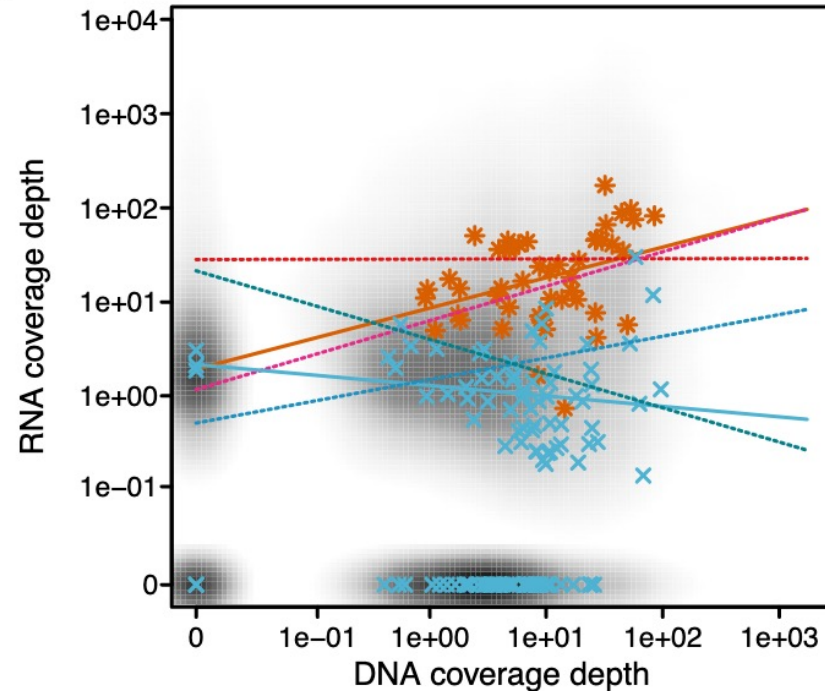
Expression per and across genomes

mOTU genes:



- motu linkage group 115 – Clostridiales
- *Roseburia inulinivorans*
- motu linkage group 316 – Clostridiales

non-house-keeping genes:

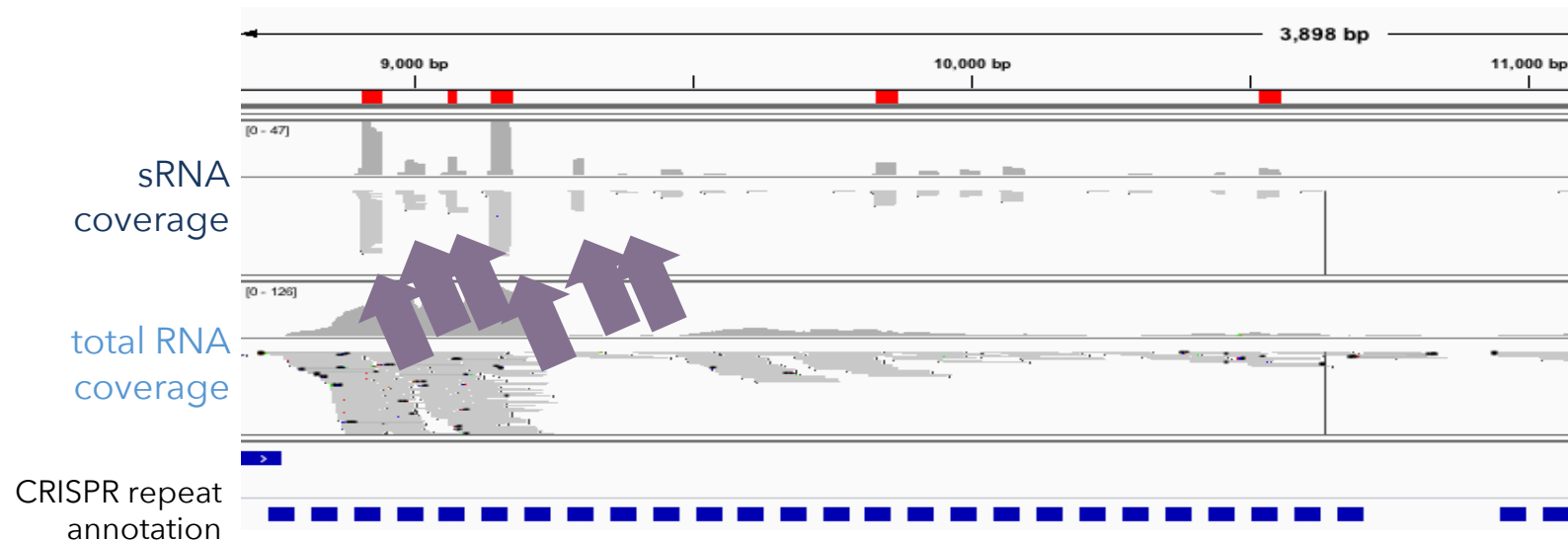


- * TIGR02013
- TIGR02013
- TIGR02350
- TIGR00981
- × KEGG:K01134
- KEGG:K01134
- KEGG:K01442
- KEGG:K01599

Example - non-coding elements in a *Parabacteroides distasonis* genome

- 4.7 Mbp
- ~4,600 ORFs
 - ~3,600 expressed
 - 16 proteins detected
- 2,949 UTRs
- 84 annotated non-coding regions (+ 4 5S rRNAs and 55 tRNAs)
- 46 CRISPR repeats
- 1 bacterial nc RNA (*Bacteroidales-1*)
- 12 riboswitches
- other potential regulatory elements

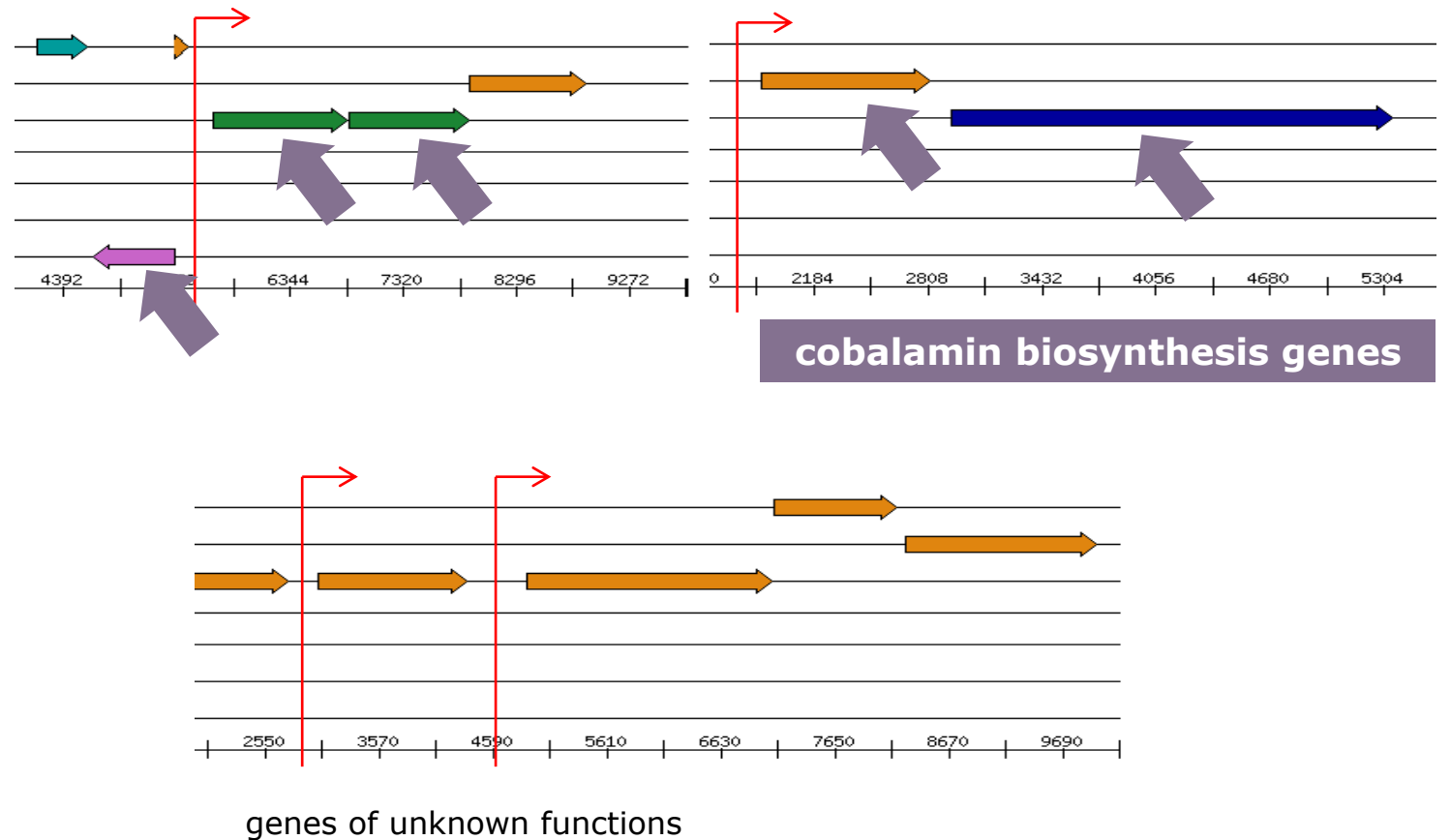
CRISPR region:



Example - non-coding elements in a *Parabacteroides distasonis* genome

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- 1 bacterial nc RNA (*Bacteroidales-1*)
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- other potential regulatory elements

cobalamin riboswitches and gene function:



Heintz-Buschart *et al.*, unpublished



Thanks for your attention!



a.u.s.heintzbuschart@uva.nl

SP C2.205



github.com/a-h-b



twitter.com/_a_h_b_